

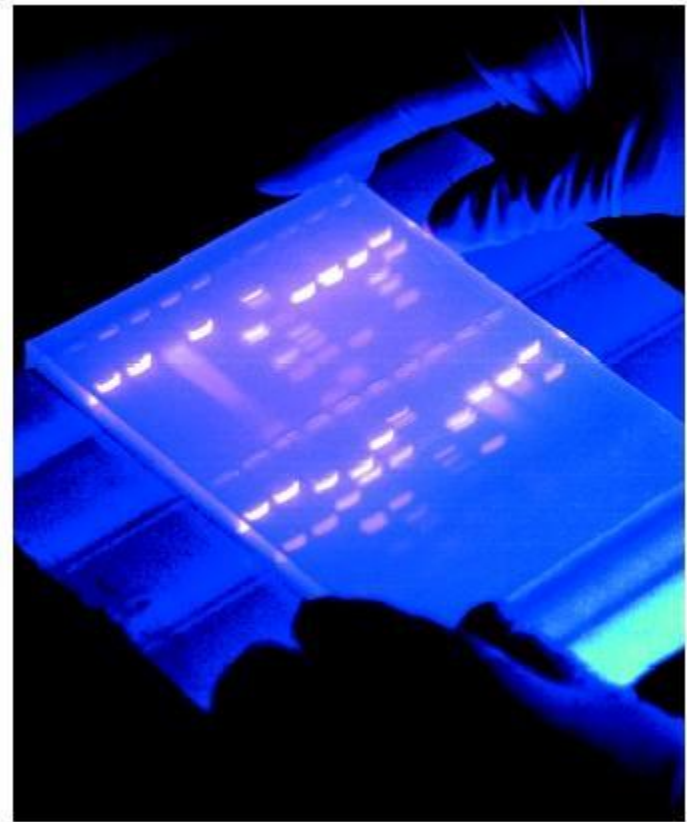
# Electrophoresis Part 2

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# Zonal Electrophoresis

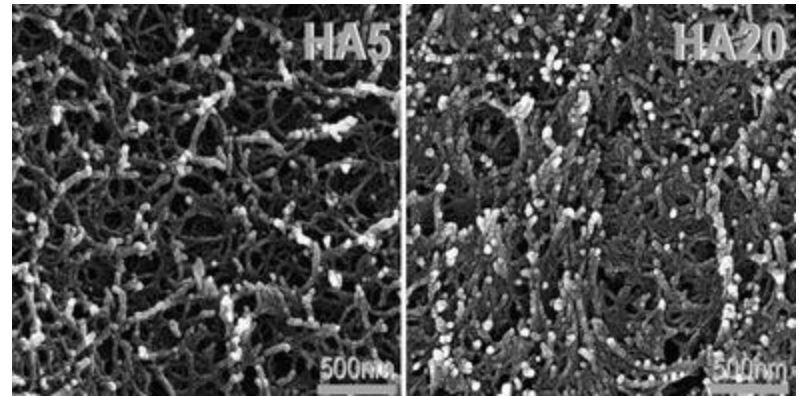
- Most common form of electrophoresis in biological studies
- Uses a support system, most commonly gel to separate proteins by their properties
- We will cover methods to separate by:
  - **Size (Through Frictional Properties)**
  - **Charge**
  - **Both**



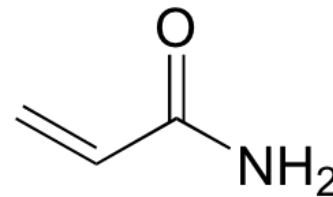
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# Gels

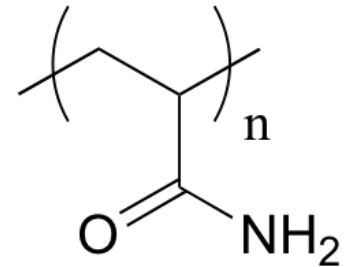
- Two main types of gels
  - **Agarose**
    - Seaweed based linear polysaccharide
    - Mechanical properties are determined by the percentage of Agarose
  - **Polyacrylamide (PAGE)**
    - Cross-linking acrylamide polymer
    - Firmness and pore size are determined by percentage of PAGE present and bisacrylamide



<http://www.rsc.org/ej/SM/2010/b926713a/b926713a-f2.gif>



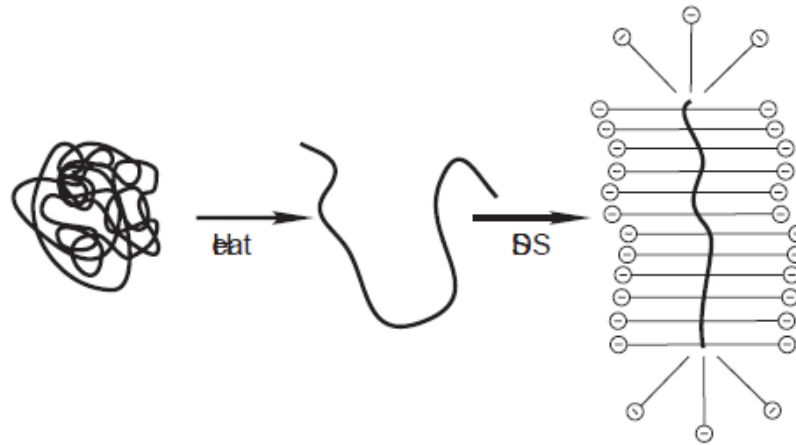
acrylamide



polyacrylamide

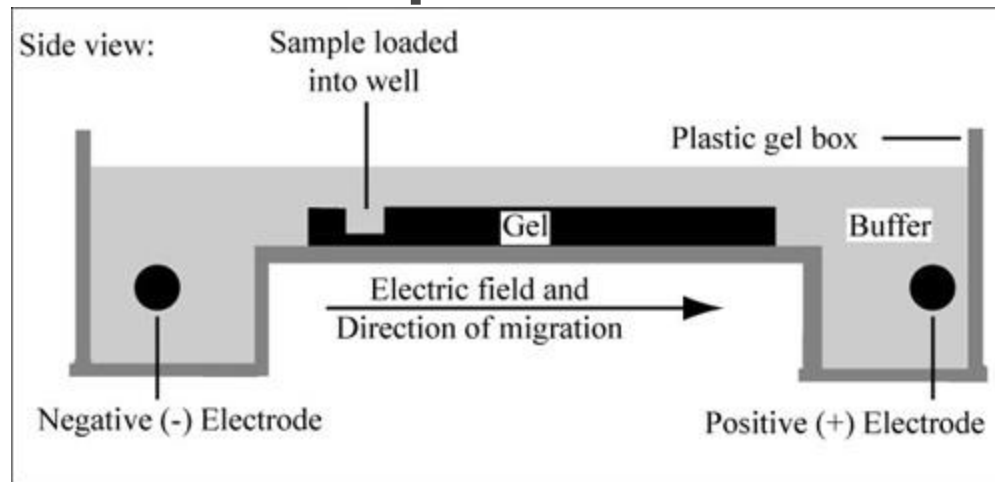
<http://chemwiki.ucdavis.edu/@api/deki/files/11311/=image084.png>

# SDS Gel Electrophoresis



- Separates proteins by size
- Proteins are denatured and negatively-charged sodium dodecyl sulfate (SDS) is added
  - SDS binds to every two amino acids causing the protein to have a negative charge
  - SDS polypeptides move through a gel at a rate dependent on their mass

# SDS Gel Electrophoresis

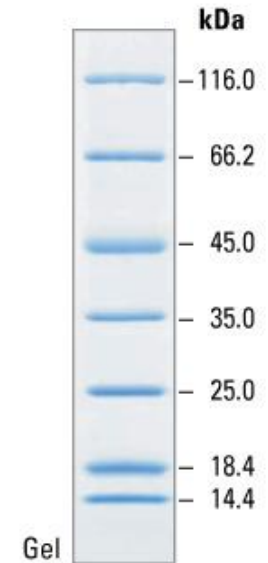
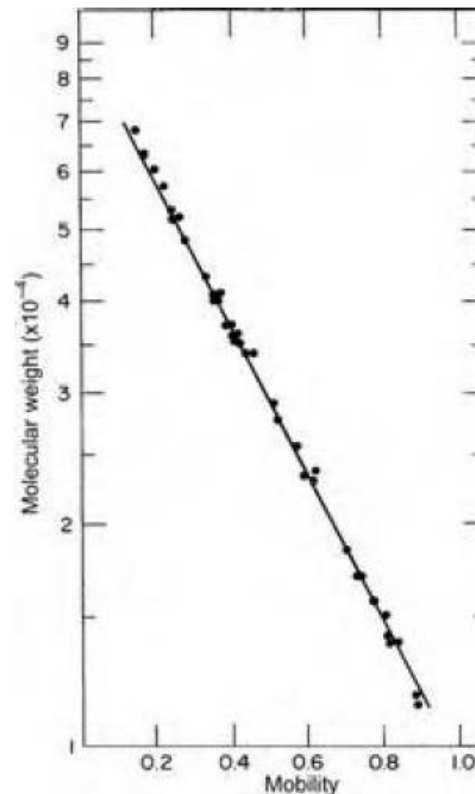


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- Protein is placed into a gel in a conductive solution and then pulled towards a positive electrode
- The distance travelled in the gel is related logarithmically to the size
- A control sample with known sizes is used to determine the sizes of the unknown samples

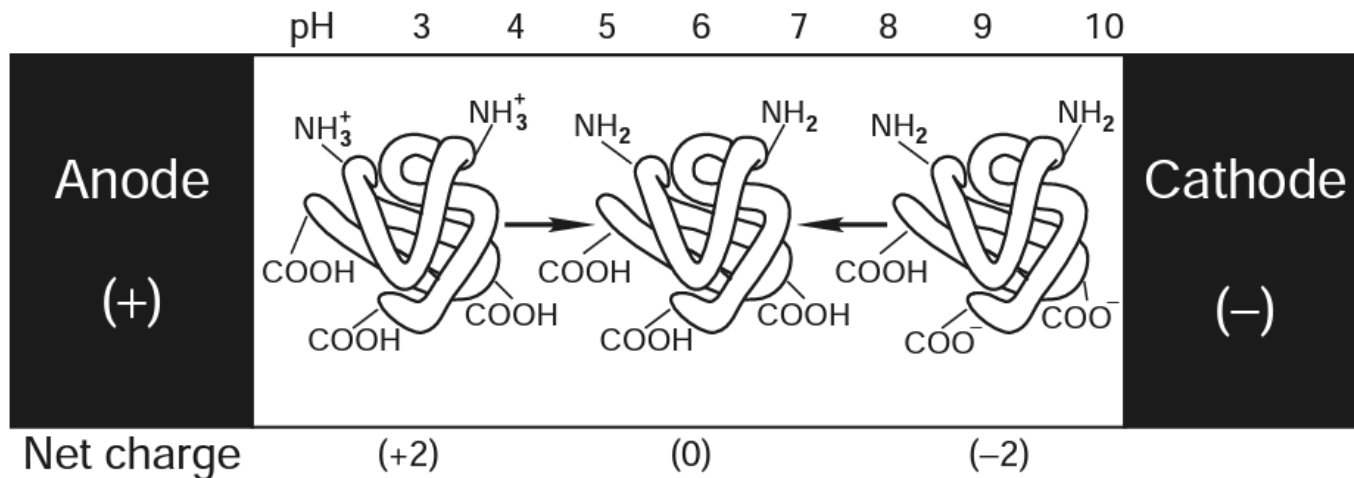
# SDS Gel Electrophoresis

- Log of the molecular weight versus the distance traveled through the gel is plotted on a semi log plot
- Known sample is used to create a line
- Unknowns are determined using the equation of the line



[http://www.thermoscientificbio.com/uploads/Images/Products/Protein\\_Electrophoresis/Protein\\_Ladders/26610-ladder-002.jpg](http://www.thermoscientificbio.com/uploads/Images/Products/Protein_Electrophoresis/Protein_Ladders/26610-ladder-002.jpg)

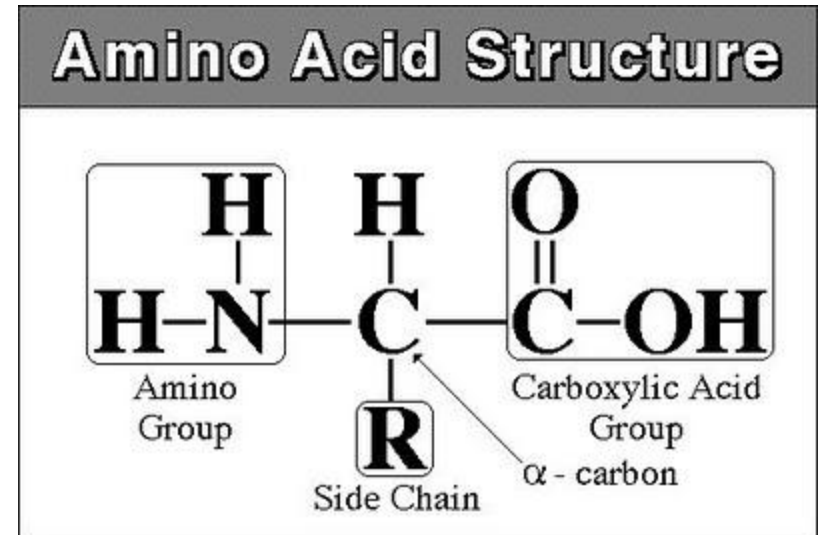
# Isoelectric Focusing (IEF)



- Separates proteins by charge
- Separates amphoteric molecules in a pH gradient
  - Amphoteric molecule are molecules whose charge is dependent on pH
  - They all have a pI point where they have neutral charge
- When a pH gradient is created across a gel, the amphoteric molecule will be pulled by the electrode until it reaches its pI point
- Once it is neutral, it is no longer affected by the electrodes

# Understanding IEF

- pKa of the carboxylic acid is around 2.2
- pKa of the amine group is 9.4
- When  $\text{pH} > \text{pKa}$ , the group is deprotonated
- When  $\text{pH} < \text{pKa}$ , the group is protonated

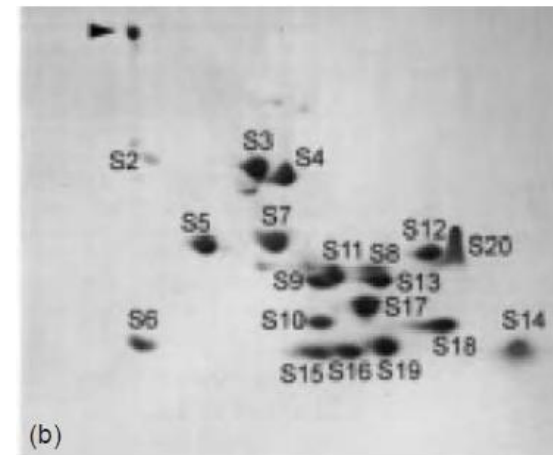
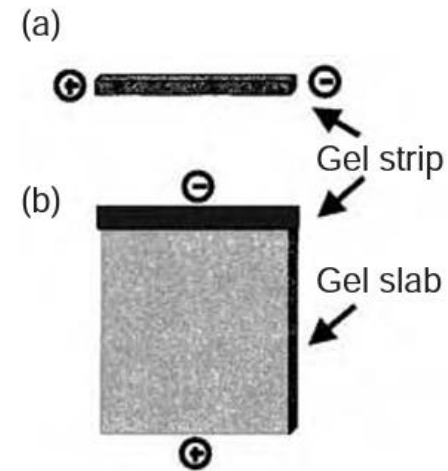


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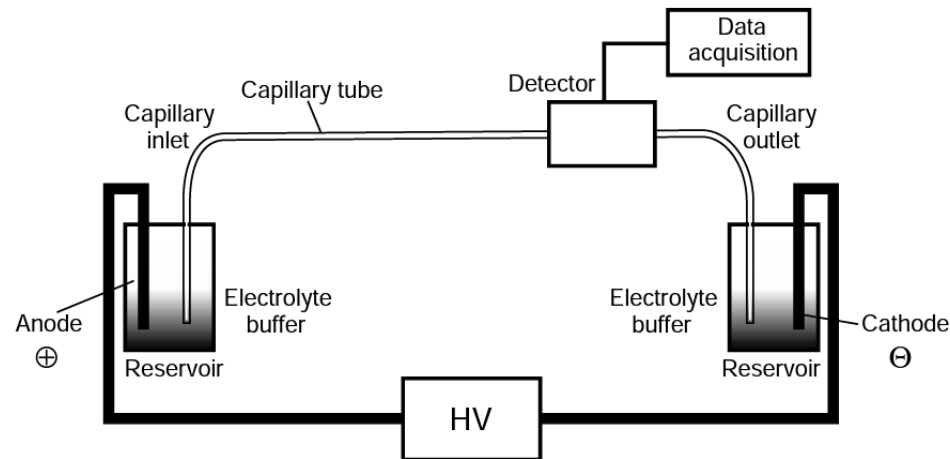


# Two-Dimensional Gel Electrophoresis

- Separates by both size and charge
- IEF is first used on a gel strip
- The strip is then mounted on a gel slab and SDS PAGE is used
- Proteins can be later removed from gel to identify
- Great for large scale comparisons (proteomics)



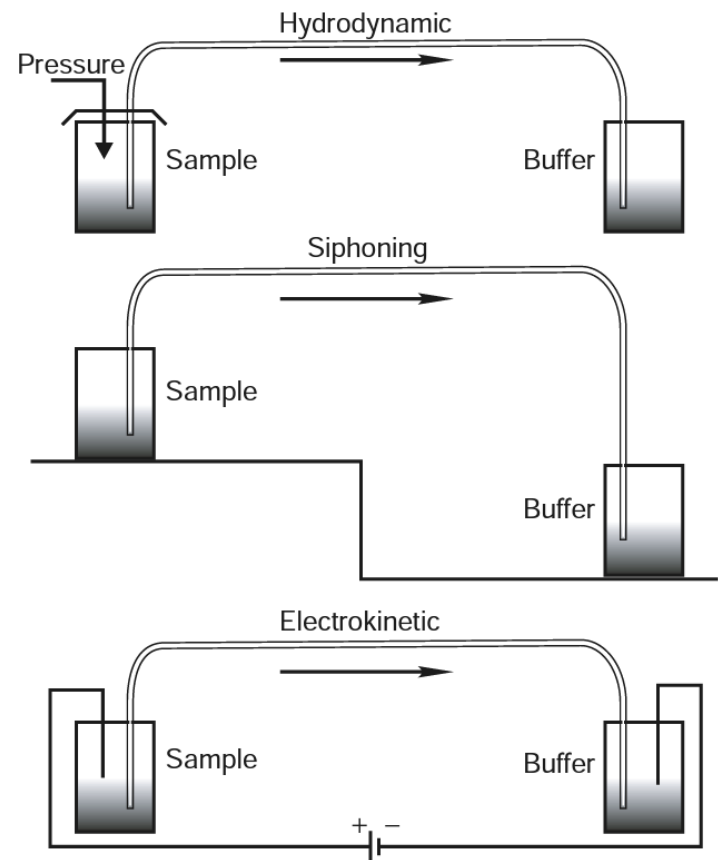
# Capillary Electrophoresis (CE)



- Alternative method to gel electrophoresis
- Proteins are dragged through a capillary tube rather than a gel
- Detector uses UV light absorbance readings to identify whether a protein separation has passed through

# CE Injection

- First the capillary tube has to be loaded with buffer
- Three methods:
  - Apply pressure or vacuum to one side
  - Use a gravity siphon
  - Drive the buffer in using a potential difference



# CE Formulas

- There are two equations for CE
- First we define the electrophoretic mobility:

$$\mu = L^2 / Vt$$

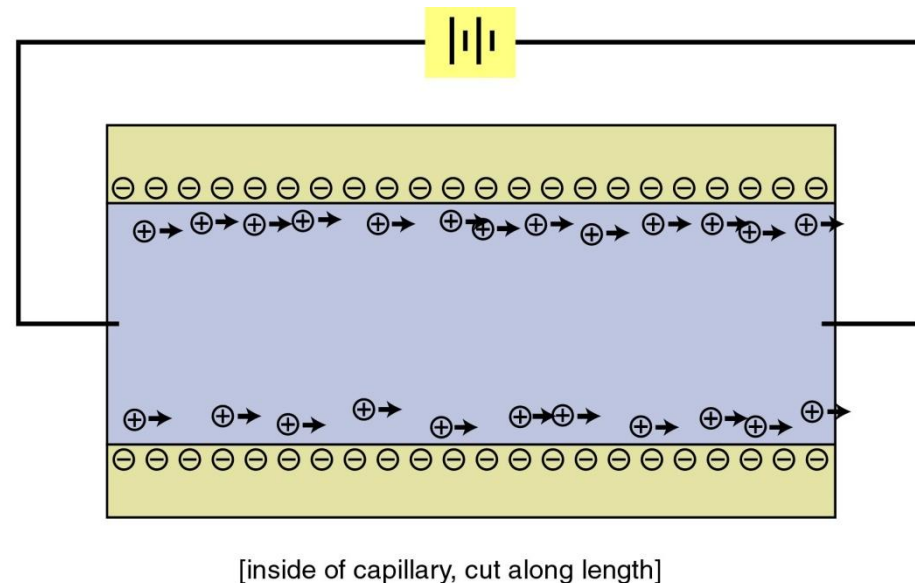
- So to minimize  $\mu$ , high voltage and short capillary are ideal
  - However we are constrained due to heat production from our power source
- Then we can look at the separation efficiency:

$$N = \mu V / 2D$$

- This is in terms of the theoretic plates, which is an evaluation of the resolution of the separation

# Electroosmotic Flow (EOF)

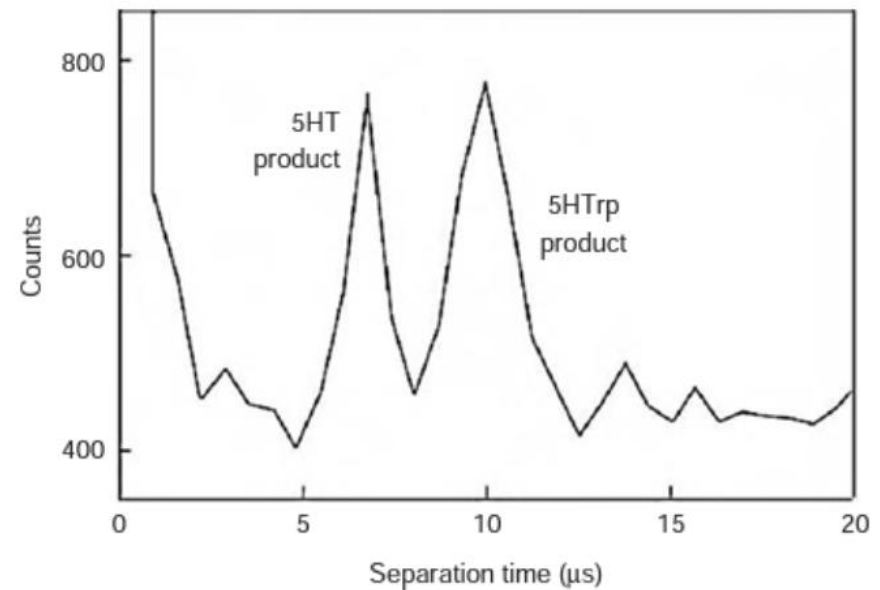
- Driving force in CE
- Drives both anions and cations towards the cathode
  - Walls of the capillary are negatively charged
  - This binds cations from the buffer
  - The cation layer that forms is attracted to the cathode and drives the flow towards the cathode
  - EOF can be reversed or completely removed by changing the charge on the capillary walls



[http://micromachine.stanford.edu/~dlaser/images/eof\\_capillary.jpg](http://micromachine.stanford.edu/~dlaser/images/eof_capillary.jpg)

# Ultrafast Capillary Electrophoresis

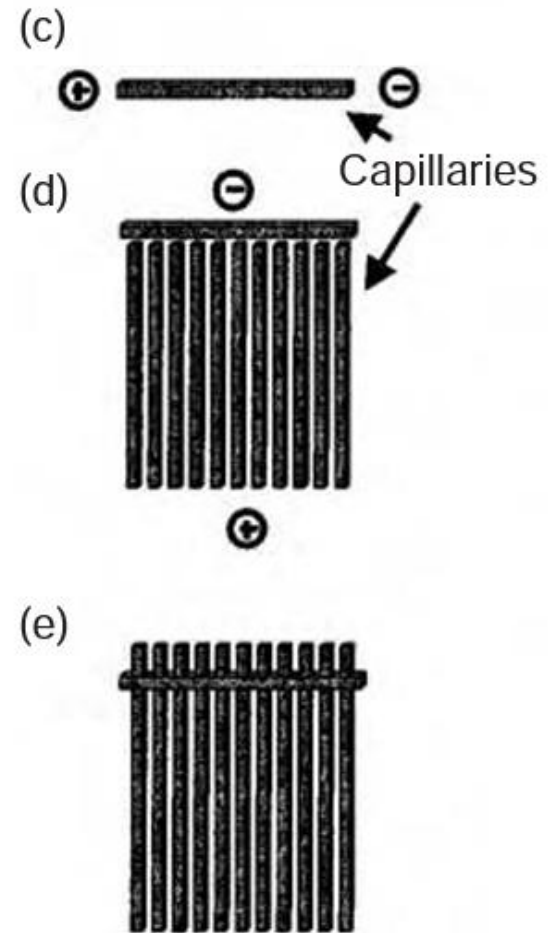
- It is possible to speed up separation
- Using an hourglass shape in the capillary increases the electric field at a specific point (reducing the risk of overheating)
- As cross-sectional area decreases electric field magnitude increases, this allows very large electric field at the mid point with relative low input voltage



**Fig. D5.17** Electrophoretic resolution of 5HT and 5HTrp photoproducts in 10  $\mu\text{s}$ . A field, which was estimated to be  $0.15 \text{ MV cm}^{-1}$  (35 kV), was used to fractionate components over a separation distance of 9  $\mu\text{m}$ . (After Plenert and Shear, 2003.)

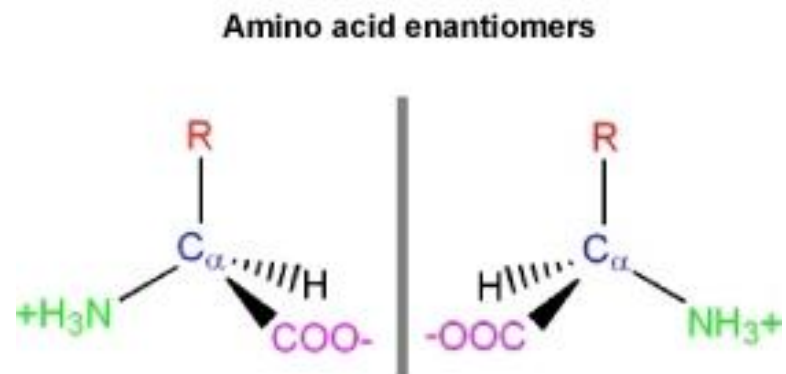
# Two-Dimensional CE

- Performed similarly to 2-D gel electrophoresis
- CE is performed in a one-dimensional capillary
- The capillary is then connected to a series of parallel capillaries
- Then a second separation is performed



# Chirality

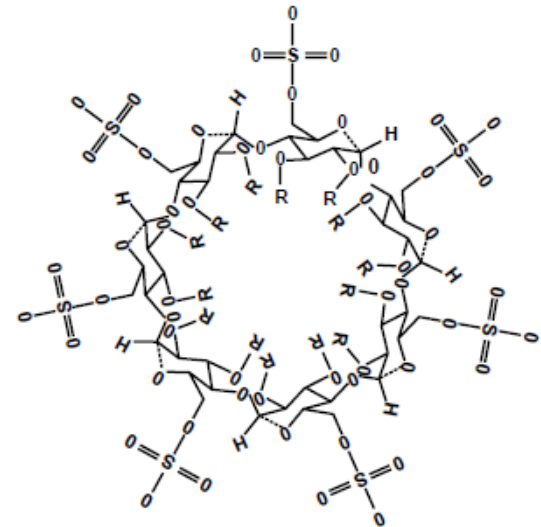
- Chirality is the “handedness” of a molecule
  - Chiral molecules are non-superimposable mirror images of each other
  - While identical in structure, chiral molecules can have vastly different properties
- Pharmaceutical applications
  - **Thalidomide**
    - One enantiomer helps with morning sickness
    - The other enantiomer causes birth defects





# Separation of Chiral Molecules

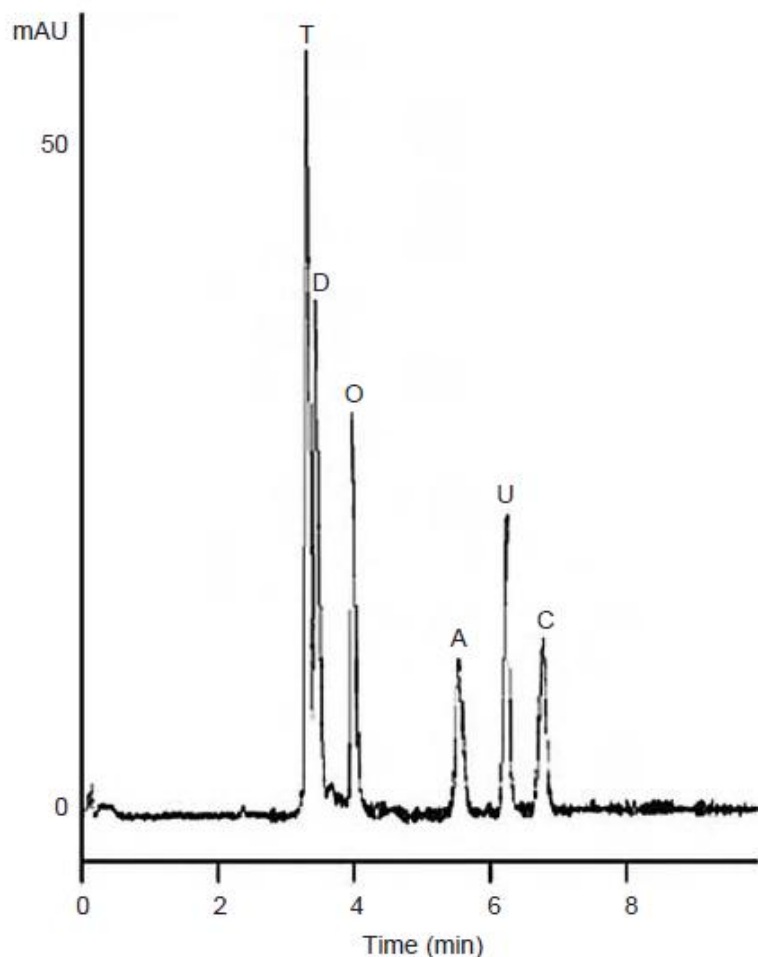
- CE is performed with Cyclodextrin in the buffer
- Cyclodextrin is a non-ionic, cyclic, chiral molecule
- One enantiomer will react much more strongly with cyclodextrin
- This enantiomer will reach the detector later
  - Interaction with the cyclodextrin slows down the migration of this enantiomer



**Fig. D5.20** Chemical structure of  $\beta$ -cyclodextrin. (After Ward, 1994.)

# Capillary Electrophoresis (CEC)

- Uses EOF to move through a column
- Sorbent in column will interact differently with different molecules
- Molecules will migrate at different rates based on their charge and based on their interactions with the sorbent in solution
- By adding an additional determining factor, it becomes easier to distinguish between molecules that would otherwise look similar (in CE)



# Sources

1. Serdyuk, Igor N., Nathan R. Zaccai, and Joseph Zaccai. *Methods in Molecular Biophysics: Structure, Dynamics, Function*. New York: Cambridge University Press, 2007. Print.
2. Voet, Donald, Judith G. Voet, and Charlotte W. Pratt. *Fundamentals of Biochemistry: Life at the Molecular Level*. 4th ed. John Wiley & Sons, Inc., 2013. Print.
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