

Static & Dynamic Light Scattering

- First quantitative experiments in 1869 by Tyndall (scattering of small particles in the air – Tyndall effect)
- 1871 – Lord Rayleigh started a quantitative study and theory
- Basic idea: incident monochromatic linearly polarized light beam incident on a sample.
Assume
 - No absorption
 - Randomly oriented and positioned scatterers
 - Isotropic scatterers
 - Independently scattering particles (dilute)
 - Particles small compared to wavelength of lightWe'll remove some of these restrictions later

Classical Wave description

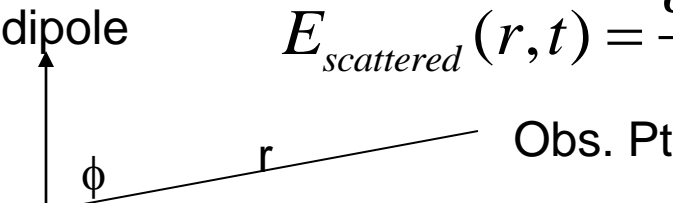
- The incident electric field is

$$E = E_0 \cos(2\pi x/\lambda - 2\pi t/T)$$

- Interaction with molecules drives their electrons at the same f to induce an oscillating dipole

$$P_{\text{induced}} = \alpha E_0 \cos(2\pi x/\lambda - 2\pi t/T) \quad - \alpha = \text{polarizability}$$

- This dipole will radiate producing a scattered E field from the single molecule


$$E_{\text{scattered}}(r, t) = \frac{\alpha E_0 4\pi^2 \sin \phi}{r \lambda^2} \cos(2\pi x / \lambda - 2\pi t / T)$$

Static (or time-average) Rayleigh scattering

1. $E \sim 1/r$ so $I \sim 1/r^2$ - necessary since $I \sim \text{energy/time/area}$ and $A \sim r^2$
2. $E \sim 1/\lambda^2$ dependence so $I \sim 1/\lambda^4$ – blue skies and red sunsets (sunrises)
3. Elastic scattering – same f
4. $\sin \phi$ dependence – when $\phi = 0$ or π – at poles of dipole – no scattering – max in horizontal plane
5. α related to n , but how?

Polarizability and index of refraction

- Note that if $n \sim 1$ $n = 1 + \frac{dn}{dc} c$
 where c is the weight concentration
- Then $n^2 = 1 + 2 \frac{dn}{dc} c + \dots$ so $n^2 - 1 = 2 \frac{dn}{dc} c = 4\pi N \alpha$
 where N = number concentration
- So, $\alpha = \frac{\frac{dn}{dc} c}{2\pi N}$ or $\alpha = \frac{M \frac{dn}{dc}}{2\pi N_A}$
- For a particle in a solvent with n_{solv} , we have $n^2 - n_{solv}^2 = 4\pi N \alpha$ so

$$\alpha = (n + n_{solv})(n - n_{solv}) / 4\pi N \sim \frac{2n_{solv}}{4\pi} \frac{dn}{dc} \frac{c}{N} = \alpha = \frac{n_{solv}}{2\pi N_A} \frac{dn}{dc} M$$

Scattered Intensity

- Detect intensity, not E, where

$$\left. \frac{I_{scatt}}{I_o} \right]_{1\text{ particle}} = \frac{\left(\frac{\alpha E_o 4\pi^2 \sin \phi}{r \lambda^2} \right)^2}{E_o^2} = \frac{16\pi^4 \alpha^2 \sin^2 \phi}{r^2 \lambda^4}$$

- Substituting for α , we have

$$\left. \frac{I_{scatt}}{I_o} \right]_{1\text{ particle}} = \frac{4\pi^2 M^2 n_{solv}^2 \left(\frac{dn}{dc} \right)^2 \sin^2 \phi}{r^2 \lambda^4 N_A^2}$$

Scattered Intensity II

- If there are N scatterers/unit volume and all are independent with $N = N_A c/M$, then

$$\left[\frac{I_{scatt}}{I_o} \right]_{per\ unit\ volume} = N \left[\frac{I_{scatt}}{I_o} \right]_{1\ particle} = \frac{4\pi^2 \sin^2 \phi n_{solv}^2 \left(\frac{dn}{dc} \right)^2 Mc}{N_A r^2 \lambda^4}$$

- We define the Rayleigh ratio R_θ :

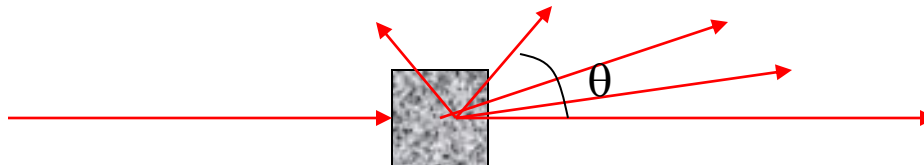
$$R_\theta = \frac{I_{scatt,\theta} r^2}{I_o \sin^2 \phi} = \frac{4\pi^2 n_{solv}^2 \left(\frac{dn}{dc} \right)^2 Mc}{N_A \lambda^4} = KMc$$

Basic Measurement

- If the intensity ratio I_{θ}/I_0 , n_{solv} , dn/dc , λ , c , ϕ , and r are all known, you can find M .
- Usually write $Kc/R_{\theta} = 1/M$
- Measurements are usually made as a function of concentration c and scattering angle θ
- The concentration dependence is given by

$$\frac{Kc}{R_{\theta}} = \frac{1}{M} + 2Bc$$

where B is called the thermodynamic virial – same as we saw before for c dependence of D (but called A)

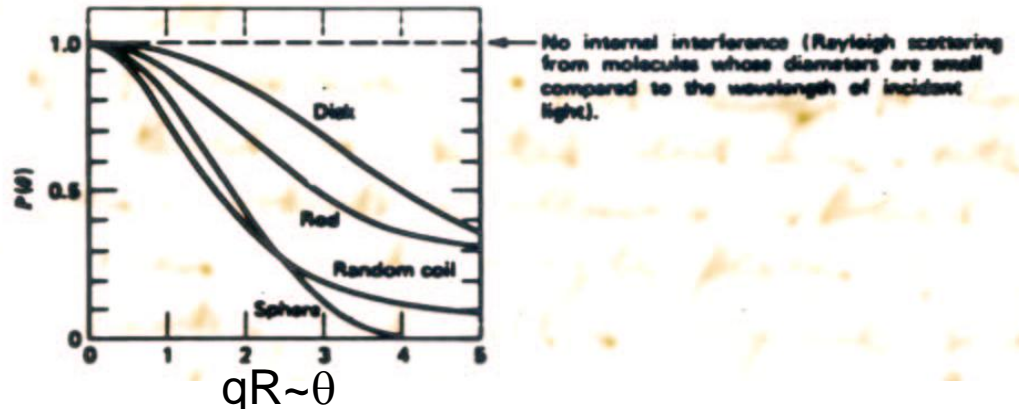


Angle Dependence

- If the scatterers are small ($d < \lambda/20$), they are called Rayleigh scatterers and the above is correct – the scattering intensity is independent of scattering angle
- If not, then there is interference from the light scattered from different parts of the single scatterer
- Different shapes give different particle scattering factors $P(\theta)$

Table 15-1 Scattering Function, $P(\theta)$, for Various Macromolecular Structures

| Structure | $P(\theta)$ | Dimensions | Radius of Gyration, R_g |
|-------------|--|--|----------------------------------|
| Sphere | $[(3a^3)^{-1}(\sin a) - a \cos a]^2$; $a = \frac{\pi D}{\lambda}$ | D = diameter of sphere | $(3/20)^{1/2} D$ |
| Thin rod | $\frac{1}{a} \int_0^a \frac{\sin x}{x} dx - \left(\frac{\sin a}{a}\right)^2$; $a = \frac{\pi D}{\lambda}$ | D = length of the rod | $D/\sqrt{3}$ |
| Random coil | $\frac{3}{a^2} [\exp(-a) + a - 1]$; $a = \frac{\pi^2 \langle D^2 \rangle}{\lambda^2}$ | $\langle D^2 \rangle$ = root-mean-square end-to-end distance | $\sqrt{\langle D^2 \rangle / 6}$ |
| Thin disk | $\frac{3}{a^2} [1 - (1/a)J_1(2a)]$; $a = (\pi D/\lambda)$ | D = diameter of disk J_1 = Bessel function order 1 | $(D/\sqrt{2})$ |

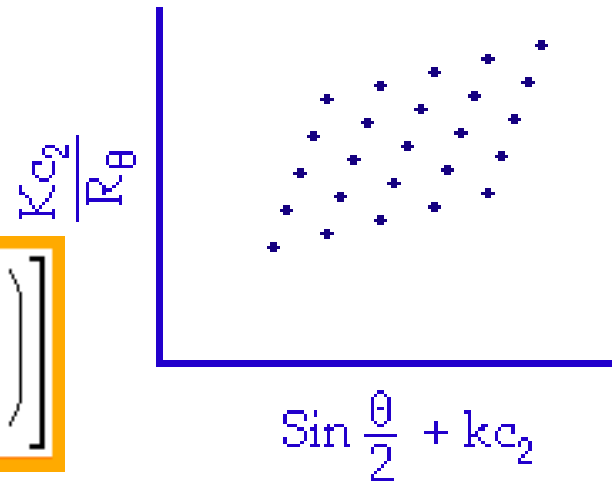


From $P(q)$, we can get a Radius of Gyration for the scatterer

Analysis of LS Data

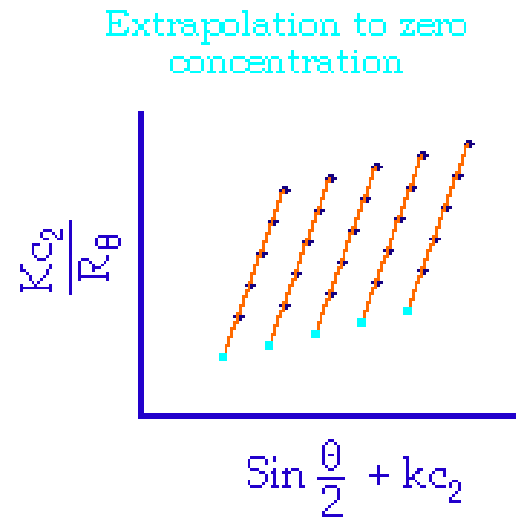
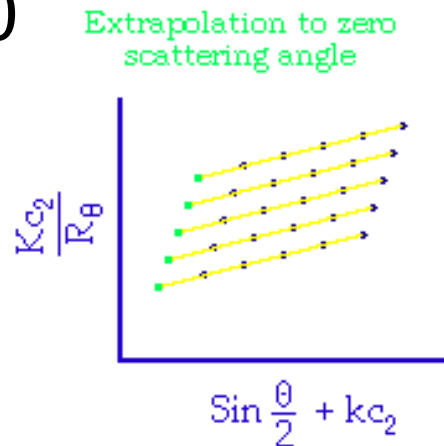
- Measure $I(\theta, c)$ and plot Kc/R_θ vs $\sin^2(\theta/2) + (\text{const})c$

$$\frac{Kc_2}{R_\theta} = \left(\frac{1}{M} + 2Bc_2 \right) \left[1 + \frac{16\pi^2 \overline{r_g^2}}{3\lambda^2} \sin^2 \left(\frac{\theta}{2} \right) \right]$$

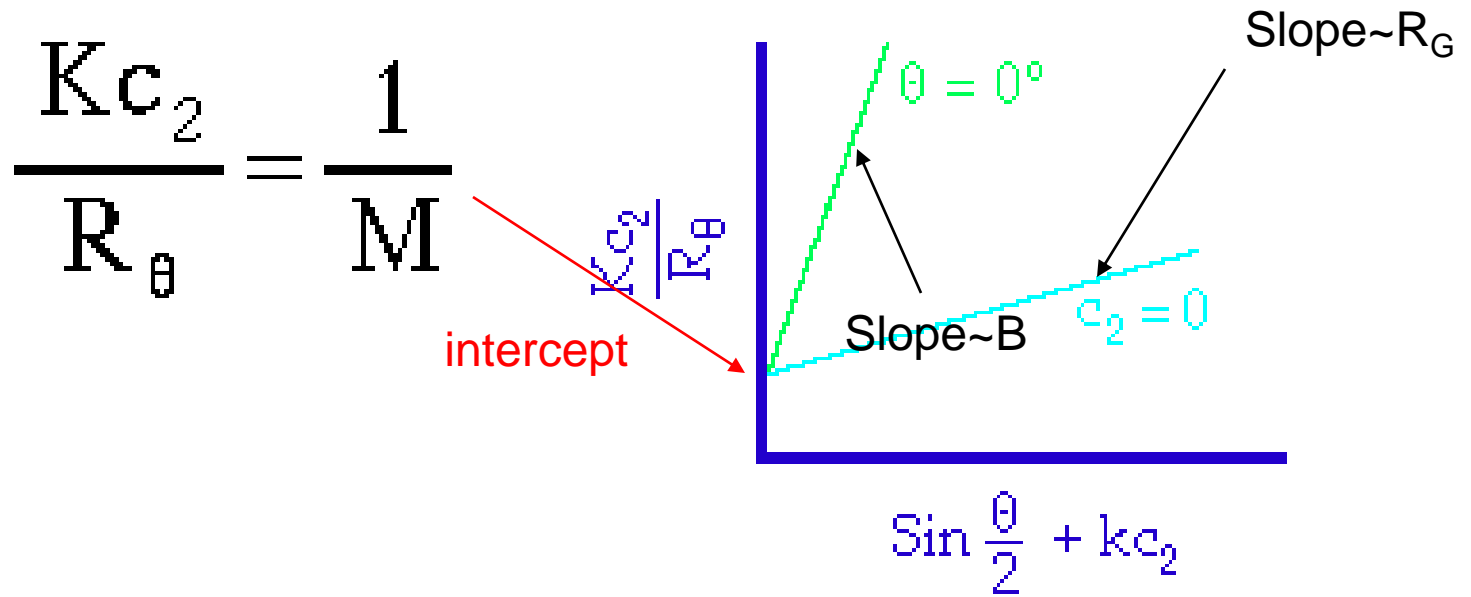


– Extrapolations: $c \longrightarrow 0$

$\theta \longrightarrow 0$



Final result



Problems: Dust, Standard to measure I_o , low angle measurement flare

Polydispersity

- If the solution is polydisperse – has a mixture of different scatterers with different M's - then we measure an average M – but which average?

$$R_{\theta} = K \frac{\sum c_i M_i}{\sum c_i} c = K \bar{M}_w c$$

- So the weight-averaged M is measured!

Possible averages:

Number-average

$$M_N = \frac{\sum M_i N_i}{\sum N_i}$$

Weight-average

$$M_w = \frac{\sum M_i c_i}{\sum c_i} = \frac{\sum M_i^2 N_i}{\sum N_i M_i}$$

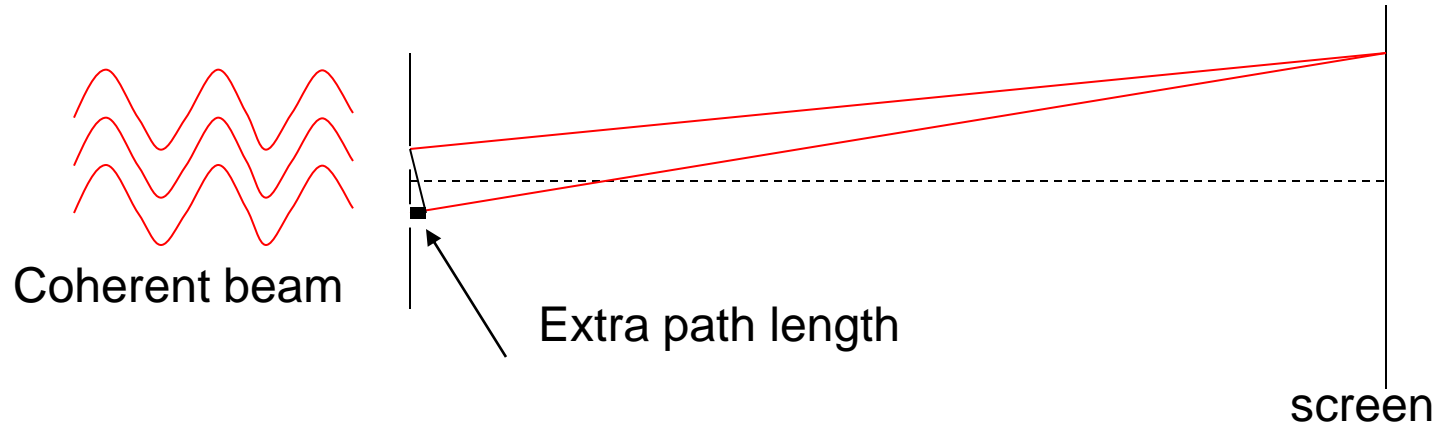
Z-average

$$M_z = \frac{\sum M_i^2 c_i}{\sum M_i c_i} = \frac{\sum M_i^3 N_i}{\sum N_i M_i^2}$$

Dynamic Light Scattering

- Basic ideas – what is it?
- The experiment – how do you do it?
- Some examples systems – why do it?

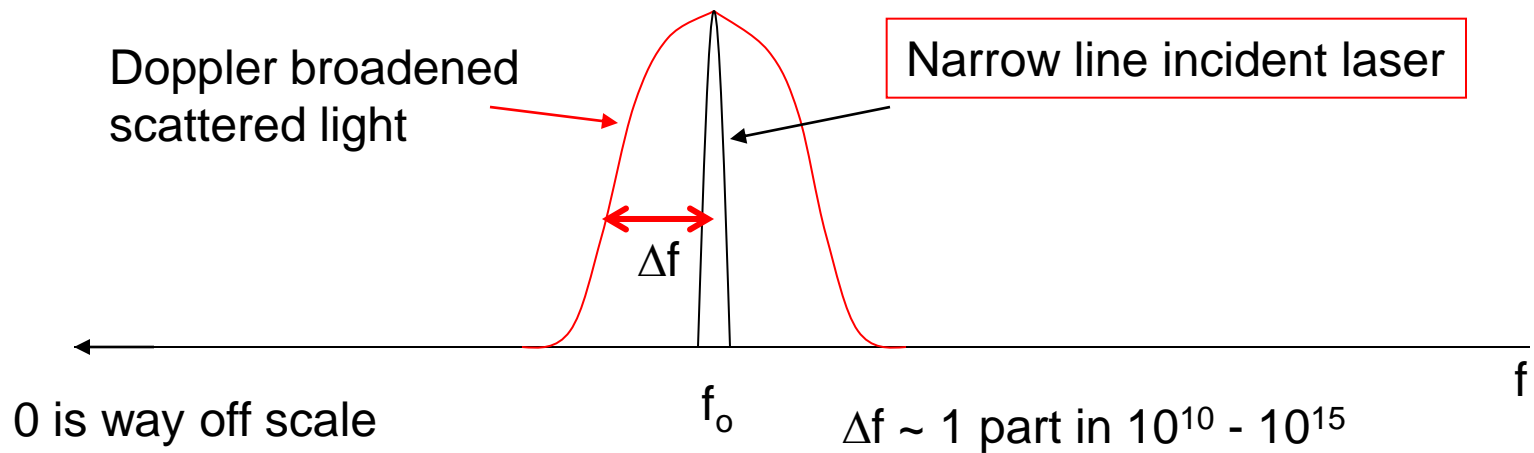
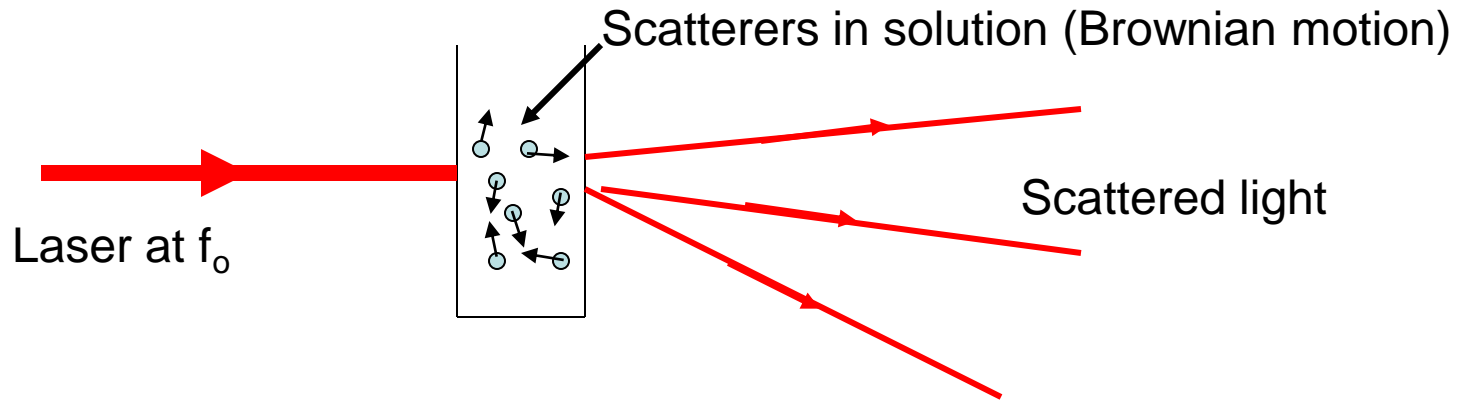
Double Slit Experiment



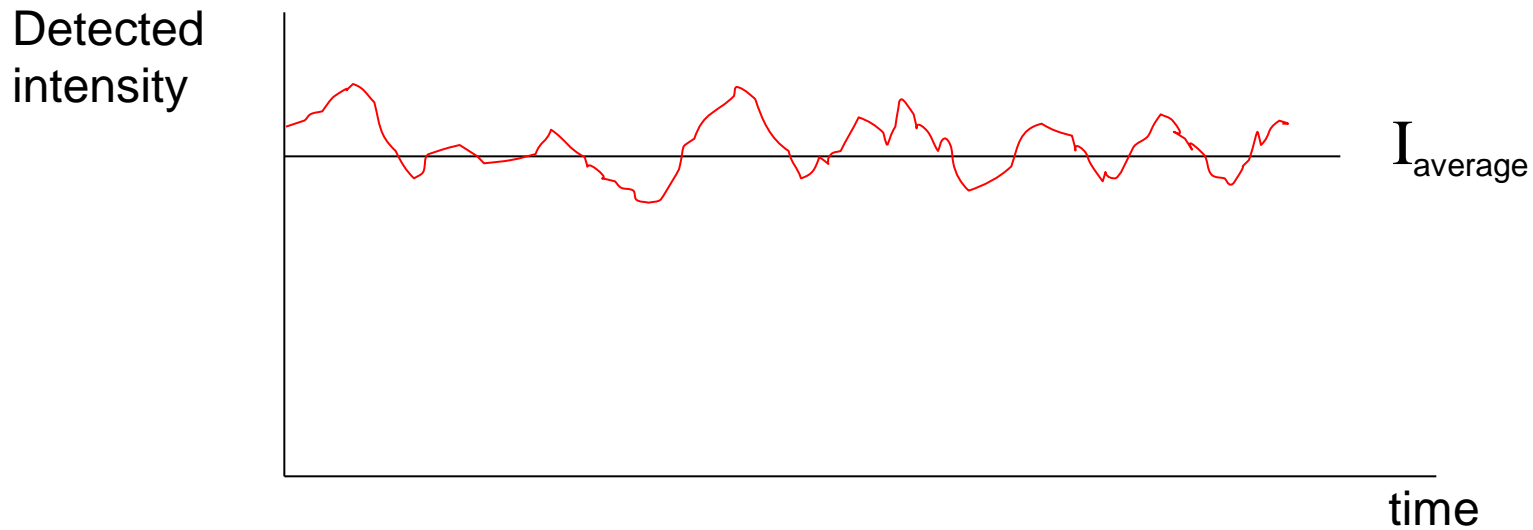
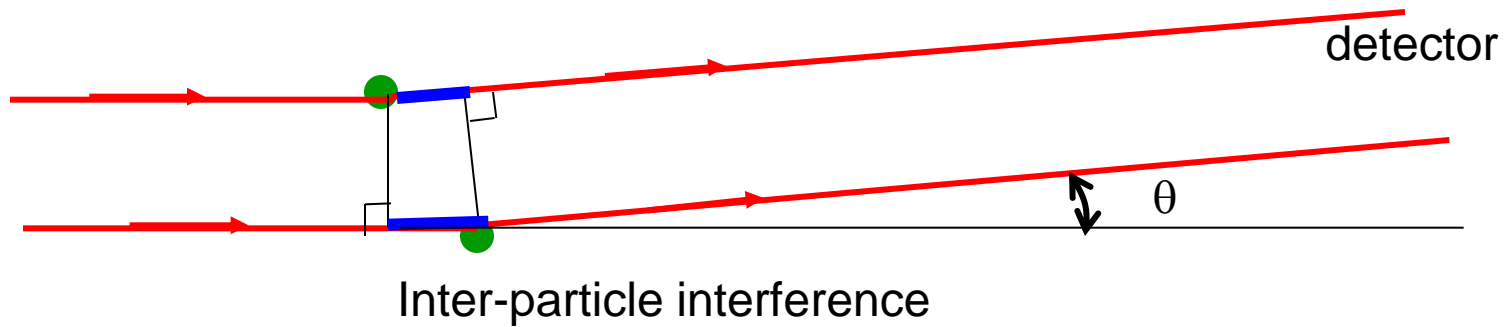
$$\begin{array}{r} + \\ \hline = \end{array} \begin{array}{c} \text{Constructive Interference} \\ \text{Two in-phase waves} \\ \text{combine to form a} \\ \text{larger wave.} \end{array}$$

$$\begin{array}{r} + \\ \hline = \end{array} \begin{array}{c} \text{Destructive Interference} \\ \text{Two out-of-phase} \\ \text{waves cancel each} \\ \text{other out.} \end{array}$$

Light Scattering Experiment



More Detailed Picture

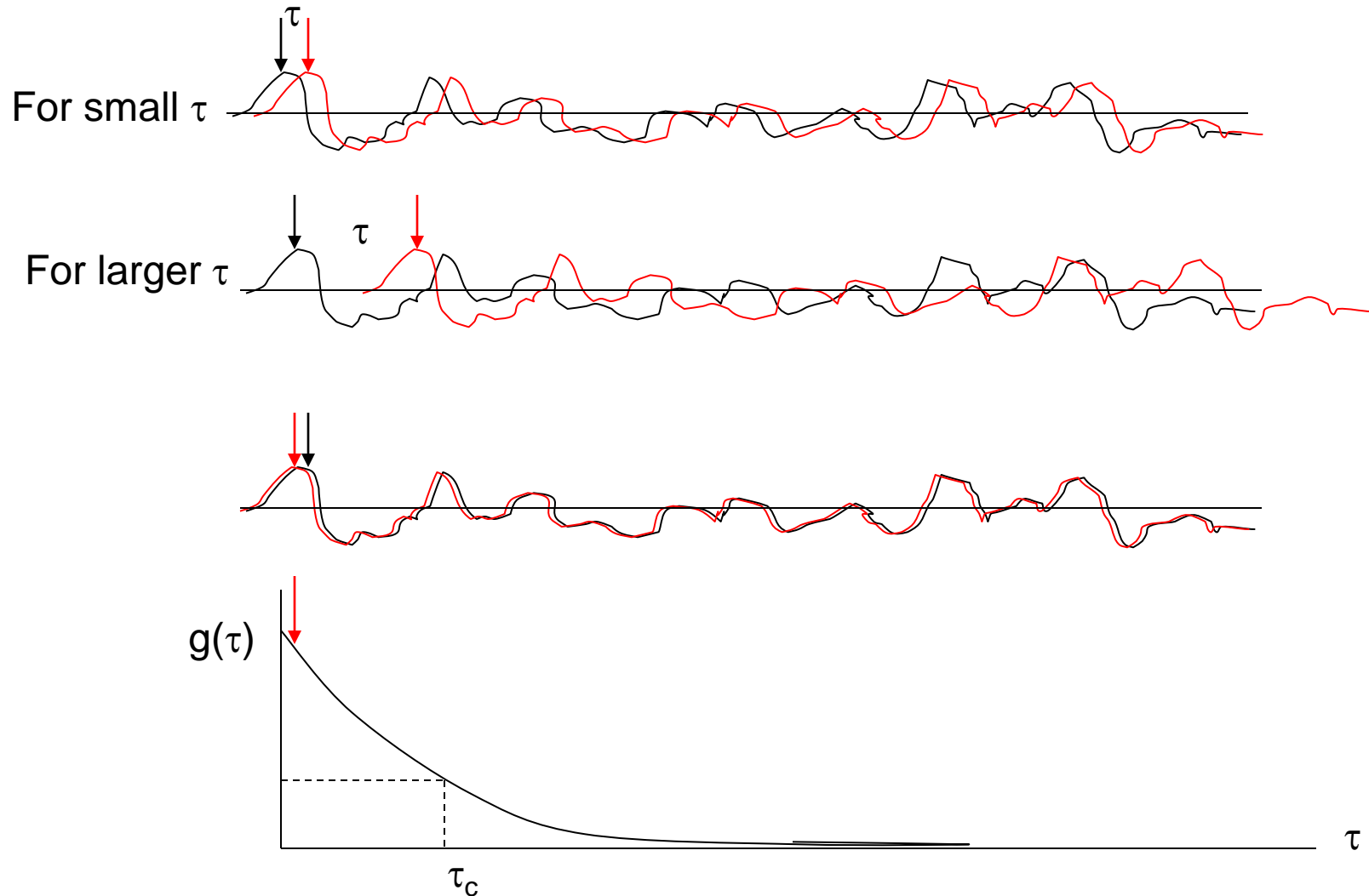


How can we analyze the fluctuations in intensity?

Data = $g(\tau) = \langle I(t) I(t + \tau) \rangle_t$ = intensity autocorrelation function

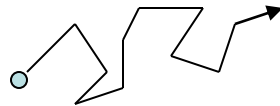
Intensity autocorrelation

- $g(\tau) = \langle I(t) I(t + \tau) \rangle_t$



What determines correlation time?

- Scatterers are diffusing – undergoing Brownian motion – with a mean square displacement given by $\langle r^2 \rangle = 6D\tau_c$ (Einstein)



- The correlation time τ_c is a measure of the time needed to diffuse a characteristic distance in solution – this distance is defined by the wavelength of light, the scattering angle and the optical properties of the solvent – ranges from 40 to 400 nm in typical systems
- Values of τ_c can range from 0.1 μs (small proteins) to days (glasses, gels)

Diffusion

- What can we learn from the correlation time?
- Knowing the characteristic distance and correlation time, we can find the **diffusion coefficient D**
- According to the Stokes-Einstein equation

$$D = \frac{k_B T}{6\pi\eta R}$$

where R is the radius of the equivalent hydrodynamic sphere and η is the viscosity of the solvent

- So, if η is known we can find R (or if R is known we can find η)

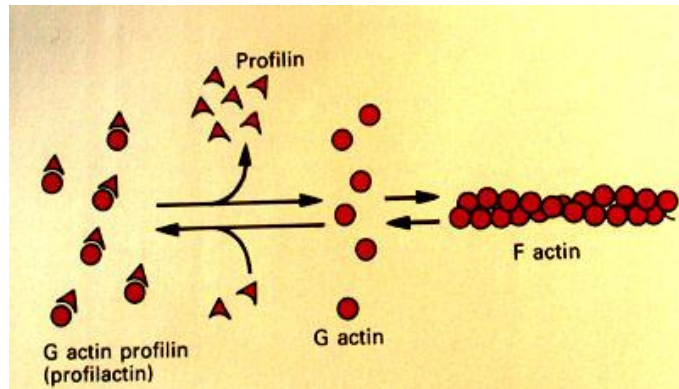
Why Laser Light Scattering?

1. Probes all motion
2. Non-perturbing
3. Fast
4. Study complex systems
5. Little sample needed

Problems: Dust and
best with monodisperse samples

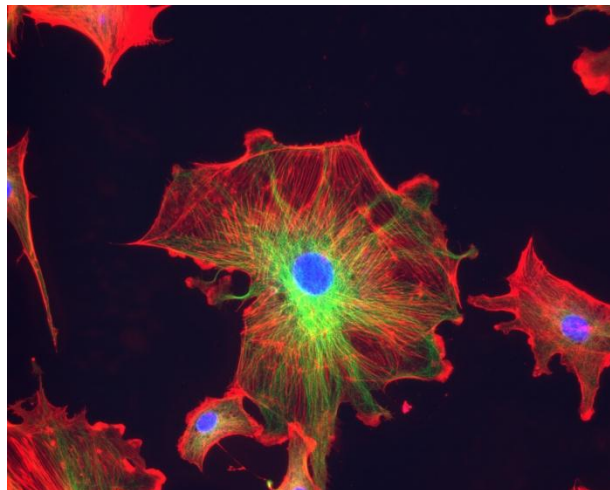
Aggregating/Gelling Systems Studied at Union College

- Proteins:
 - Actin – monomers to polymers and networks



Study monomer size/shape,
polymerization kinetics,
gel/network structures formed,
interactions with other actin-binding
proteins

Why?



Epithelial cell under fluorescent
microscope

Actin = red, microtubules = green,
nucleus = blue

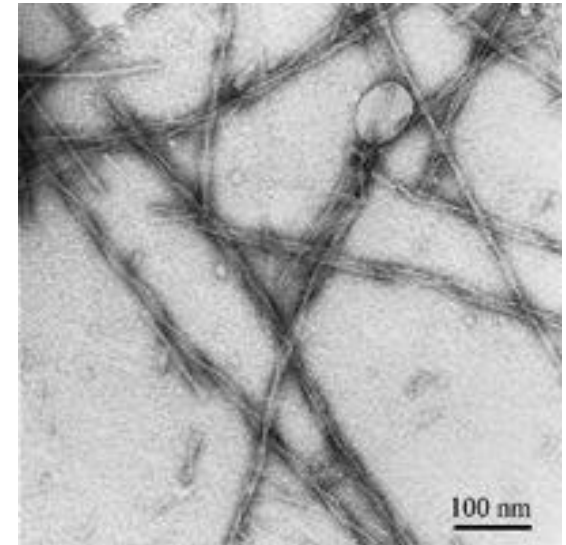
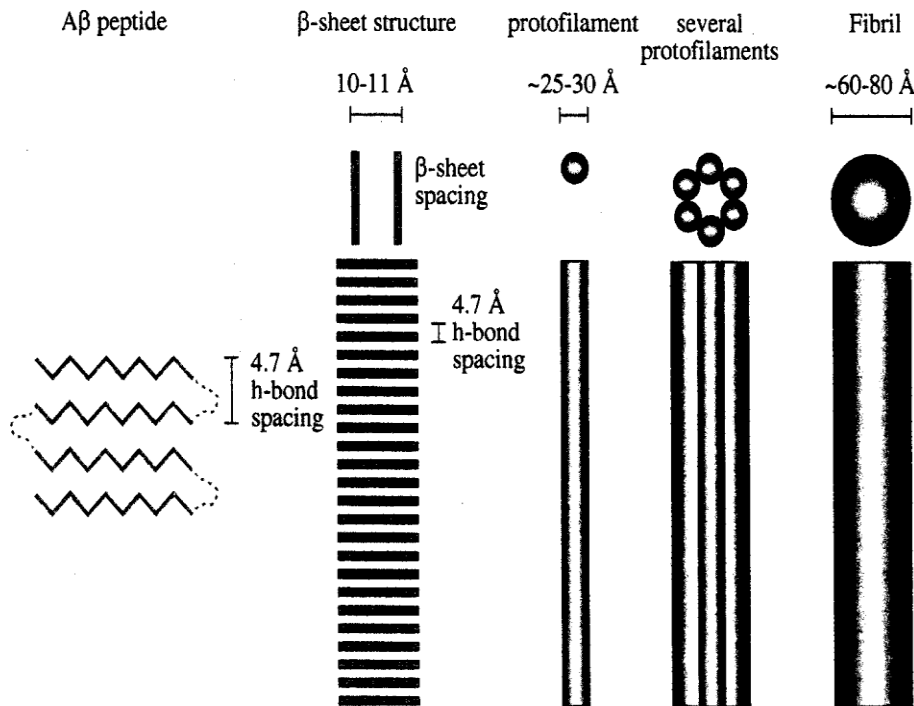
Aggregating systems, con't

- BSA (bovine serum albumin) what factors cause or promote aggregation?
- beta-amyloid +/- chaperones how can proteins be protected from aggregating?
- insulin what are the kinetics?

- Polysaccharides: Focus on the onset of gelation –
 - Agarose what are the mechanisms causing gelation?
 - Carageenan how can we control them?
what leads to the irreversibility of gelation?

Current Projects

1. **β -amyloid** – small peptide that aggregates in the brain – believed to cause Alzheimer's disease-



Current Projects

2. Insulin aggregation

EFFECTS OF ARGININE ON THE KINETICS OF BOVINE INSULIN AGGREGATION STUDIED BY DYNAMIC LIGHT SCATTERING

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